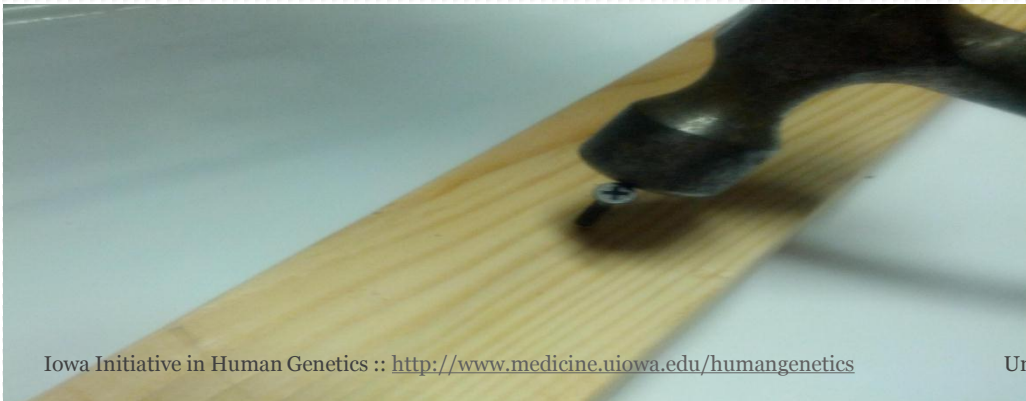


Other applications of MPS

Appropriate uses for MPS



Tom Bair
thomas-bair@uiowa.edu

Types of workflows

- reduce
 - assemble
 - align
- identify significant areas:
 - peak-finding/depth
 - snp calling
- Annotate areas of interest
 - Coordinates -> information
- display
 - Understand the information

Data reduction

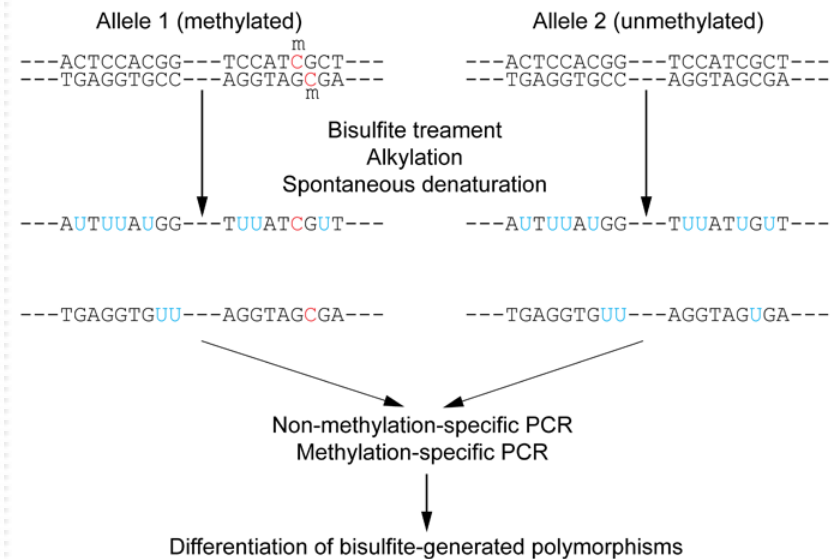
- Alignment
 - Bowtie, bwa ...
- Assemble
 - Velvet, Mira ...
- Some small genome assembly
 - High memory requirements
 - Probably will shift as longer reads are economical
 - Source data critical
 - Long paired end or just long reads make everything easy.



Identification of significant areas

- SNP/deletion discovery
 - --the focus of this course
- Chip-seq
 - Find DNA protein interactions
 - Depth of alignment vs background
- RNA-seq
 - Find differentially expressed transcripts
 - Depth of one sample vs another
 - Linkage of areas
 - Gene definition

- Methylation patterns
 - Somewhat unique alignment protocol
 - Look at the alignment of a couple different genome versions to see which matches the best, this gives the methylation status of the area
 - Bismark does this but downstream outputs limited



- Metagenomics
 - Align to a specific database
 - Greengenes or similar of rRNA sequences
 - Align to database of all sequences
 - Find distribution of genus/species

Display approaches

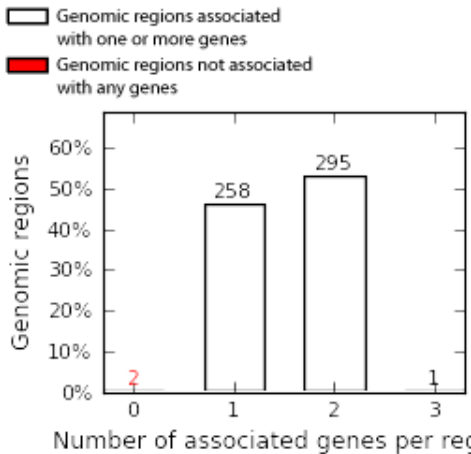
- Level of detail
- IGV genome browser
 - High level of detail, can potentially see every read, local so annotation may lag
- UCSC genome browser
 - Difficult to get all your read information uploaded
 - Very current annotation
- Trackster
 - Galaxy specific, developing

Chip Seq workflow

- Sample prep and antibody critical
- Align with bowtie2
- Peak calling with MACs
 - Mosaics – deeper sequenced samples
 - Dbchip – merge experiments
- Annotation
 - Cistrome
 - GREAT
 - Typically need adjacent genes and distance to TSS

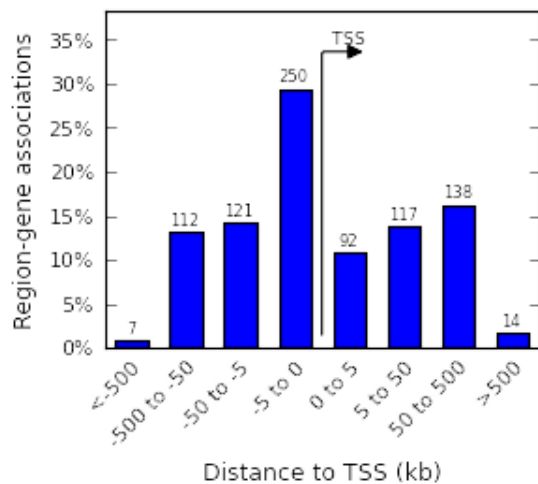
Number of associated genes per region

[Download as PDF.](#)



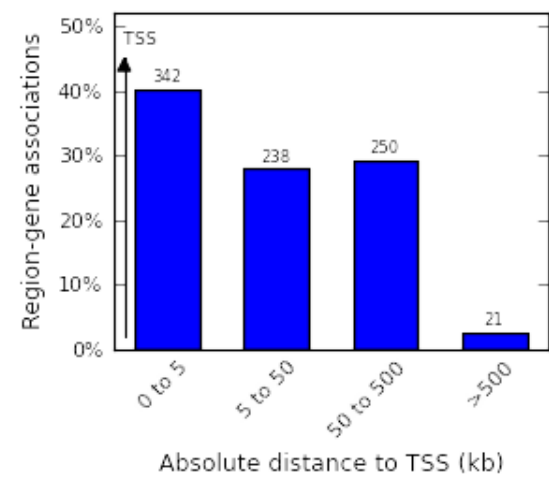
Binned by orientation and distance to TSS

[Download as PDF.](#)



Binned by absolute distance to TSS

[Download as PDF.](#)



GO Cellular Component (5 terms) Global controls

Table controls: Shown top rows in this table: Term annotation count: Min: Max: Visualize this table:

| Term Name | Binom Rank | Binom Raw P-Value | Binom FDR Q-Val | Binom Fold Enrichment | Binom Observed Region Hits | Binom Region Set Coverage | Hyper Rank | Hyper FDR Q-Val | Hyper Fold Enrichment | Hyper Observed Gene Hits | Hyper Total Genes | Hyper Gene Set Coverage |
|-----------------------|------------|-------------------|-----------------|-----------------------|----------------------------|---------------------------|------------|-----------------|-----------------------|--------------------------|-------------------|-------------------------|
| actin cytoskeleton | 1 | 9.3775e-11 | 1.0512e-7 | 2.8281 | 50 | 8.99% | 2 | 1.1912e-5 | 2.6396 | 40 | 337 | 5.04% |
| stress fiber | 2 | 1.5680e-7 | 8.7888e-5 | 6.6040 | 13 | 2.34% | 22 | 1.7569e-2 | 4.4477 | 8 | 40 | 1.01% |
| actin filament bundle | 5 | 2.5918e-7 | 5.8107e-5 | 6.3139 | 13 | 2.34% | 23 | 2.8131e-2 | 4.1374 | 8 | 43 | 1.01% |
| actomyosin | 13 | 3.1716e-6 | 2.7349e-4 | 5.0198 | 13 | 2.34% | 21 | 1.8282e-2 | 4.0029 | 9 | 50 | 1.13% |
| ruffle | 33 | 1.4147e-4 | 4.8056e-3 | 2.7582 | 18 | 3.24% | 16 | 9.2660e-3 | 3.0052 | 15 | 111 | 1.89% |

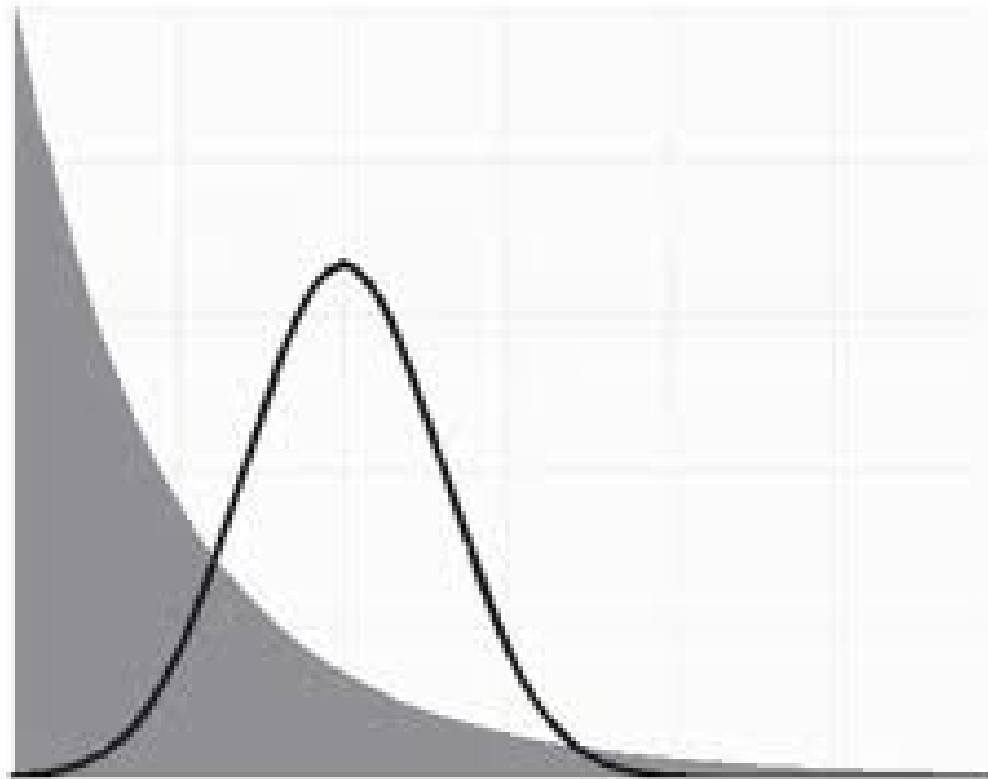
Exon re-sequencing

- BWA alignment
- GATK calibration and SNP identification
- Annotation SIFT, SVA, ANNOVAR
- IGV for visualization
- Trending to genomic rather than exon capture
 - Still may filter down to subset
 - Less bias for CNV
 - Cost, capture designed to reduce sequencing but sequencing dropping in cost
 - Still have dark spots

RNA-Seq Workflow

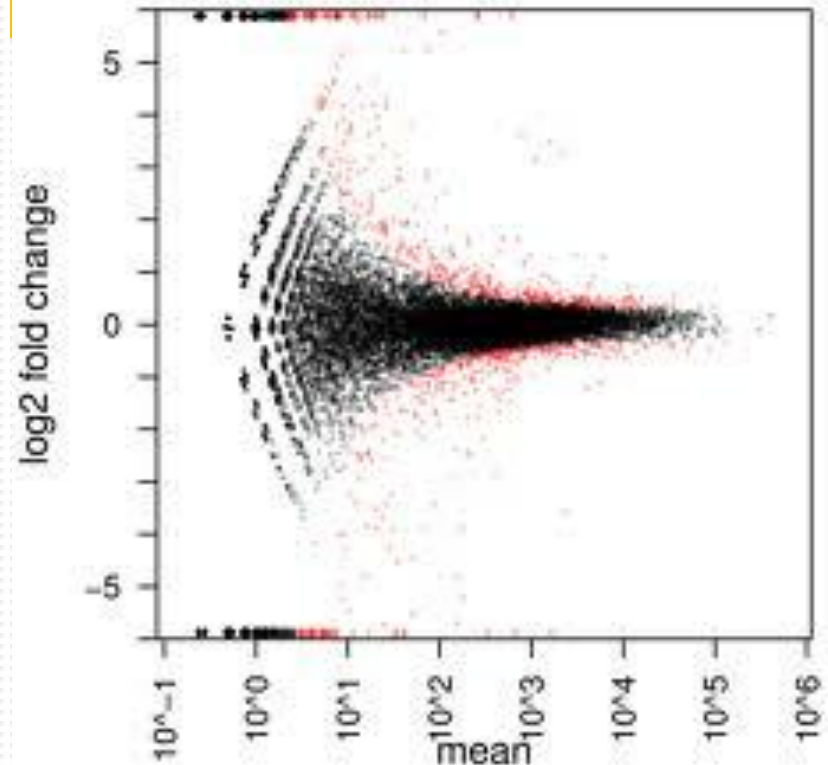
- Tophat alignment
- Htseq-count or cufflinks for counts
- EdgeR, DESeq, or cuffdiff for differential expression
 - ANOVA, t-test do not work well

Distributions



RNA-Seq length bias

- It is easier (higher statistical power) to find changes in genes that have high expression
- You may find that pathways predicted by microarray tools are simply composed of high numbers of long genes
- This is a caution that applies to many MPS techniques that have a similar array based technique.



RNA-seq w/o Reference

- Trinity
 - 16G of RAM minimum start
 - More is better
 - Active community and optimization
 - Easy paths to differential expression
 - Easy paths to ORF characterization

Methylation

- Specific alignment requirements
 - Use bowtie with specialized references
- Differential modification a similar problem to RNA-Seq
- Annotation similar to CHIP-Seq'
- Bismark a popular option for Display as well as analysis

Metagenomics

- Longer reads make everything easier/more accurate
- First data reduction typically a clustering to merge identical reads
 - The clusters are then assigned an OTU (Operational taxonomic units)

- QIIME - nice tool to assign and display output
- Krona – nice display tool



Local resources

- Galaxy
 - Harness local compute cluster helium with web page ease and interface
- Partek
 - Statistical analysis package
- Genego
 - Pathway annotation and mining tool
- Laser gene- NGS module may be soon
 - <http://cs.its.uiowa.edu/software/lasergene.shtml>
- IIHG bioinformatics core
 - <http://bioinform-div.healthcare.uiowa.edu/>

Partek

Partek Genomics Suite - Version 6.12.0531 - 11 (FinalReport_AVGSignal.txt)

File Edit Transform View Stat Filter Tools Window Custom Help

1 (FinalReport_AVGSignal.txt)
 1 (ANOVAResults.csv) *
 2 (pvalue_05_fc_2.csv)

2 (FinalReport_AVGSignal.txt)
 ANOVA-1way (ANOVAResults.csv)
 ANOVA-1way:2 (ANOVAResults-18-removed.csv)
 Ecsod_vs_Empty (Ecsod vs. Empty All.csv)
 Ecsod_vs_Empty1 (Ecsod vs. Empty_18_removed.csv)

3 (heatmap_18Anova_subset.txt)

4 (FinalReport_AVGSignal.txt) *
 ANOVA-1way (wt_v_ko.csv)
 ANOVA-1way:2 (fold-change-comparisons.csv)
 ANOVA-1way:3 (wt_v_ko_nopmn.csv)

5 (ptmp9) *

6 (FinalReport_AVGSignal.txt)

7 (heatmap_subset.txt)

8 (FinalReport_AVGSignal.txt)

9 (6-26-12_data.csv) *

10 (gse3013) *

11 (FinalReport_AVGSignal.txt)
 ANOVA-1way (ANOVAResults) *

Current Selection KD002

| 1. Sample ID | 2. Gender | 3. Index | 4. Sample Group | 5. Sentrix Barcode | 6. Sample Section | 7. Detected Genes (0.01) | 8. Detected Genes (0.05) | 9. Signal Average | 10. Signal P05 | 11. Signal P25 | 12. Signal SD |
|--------------|-----------|----------|-----------------|--------------------|-------------------|--------------------------|--------------------------|-------------------|----------------|----------------|---------------|
| 2. KD002 | ? | 2 | Group 1 | 8363851022 | B | 10245 | 12660 | 380.922 | -10.5054 | -0.816138 | 1 |
| 3. KD004 | ? | 3 | Group 1 | 8363851022 | C | 11262 | 14105 | 381.789 | -10.7404 | -0.948744 | 1 |
| 4. KD014 | ? | 4 | Group 2 | 8363851022 | D | 11349 | 14222 | 380.539 | -10.5505 | -0.912198 | 1 |
| 5. KD015 | ? | 5 | Group 2 | 8363851022 | E | 11132 | 13690 | 383.023 | -10.7708 | -0.935757 | 1 |
| 6. KD016 | ? | 6 | Group 2 | 8363851022 | F | 10860 | 14083 | 384.322 | -10.7618 | -0.896563 | 1 |
| 7. KD027 | ? | 7 | Group 3 | 8363851022 | G | 11025 | 13670 | 382.813 | -10.601 | -0.936464 | 1 |
| 8. KD028 | ? | 8 | Group 3 | 8363851022 | H | 11243 | 14305 | 382.328 | -10.5783 | -0.96665 | 1 |
| 9. KD029 | ? | 9 | Group 3 | 8363851022 | I | 10020 | 13026 | 388.432 | -10.6436 | -0.7854 | 1 |
| 10. KD0038 | ? | 10 | Group 4 | 8363851022 | J | 11338 | 14388 | 380.777 | -10.5948 | -0.920542 | 1 |
| 11. KD0039 | ? | 11 | Group 4 | 8363851022 | K | 11187 | 13959 | 384.446 | -10.746 | -0.952128 | 1 |
| 12. KD0040 | ? | 12 | Group 4 | 8363851022 | L | 11223 | 13612 | 381.899 | -10.7233 | -0.958697 | 1 |

Workflows Gene Expression

Gene Expression

Import

- Import samples
- Add sample attributes
- Edit sample information
- Choose sample ID column

QA/QC

- QC metrics
- Plot sample histogram ✓
- Principal components analysis (PCA) ✓

Analysis

- Detect differentially expressed genes ✓
- Plot sources of variation
- Create gene list
- Order TaqMan® Assays

Visualization

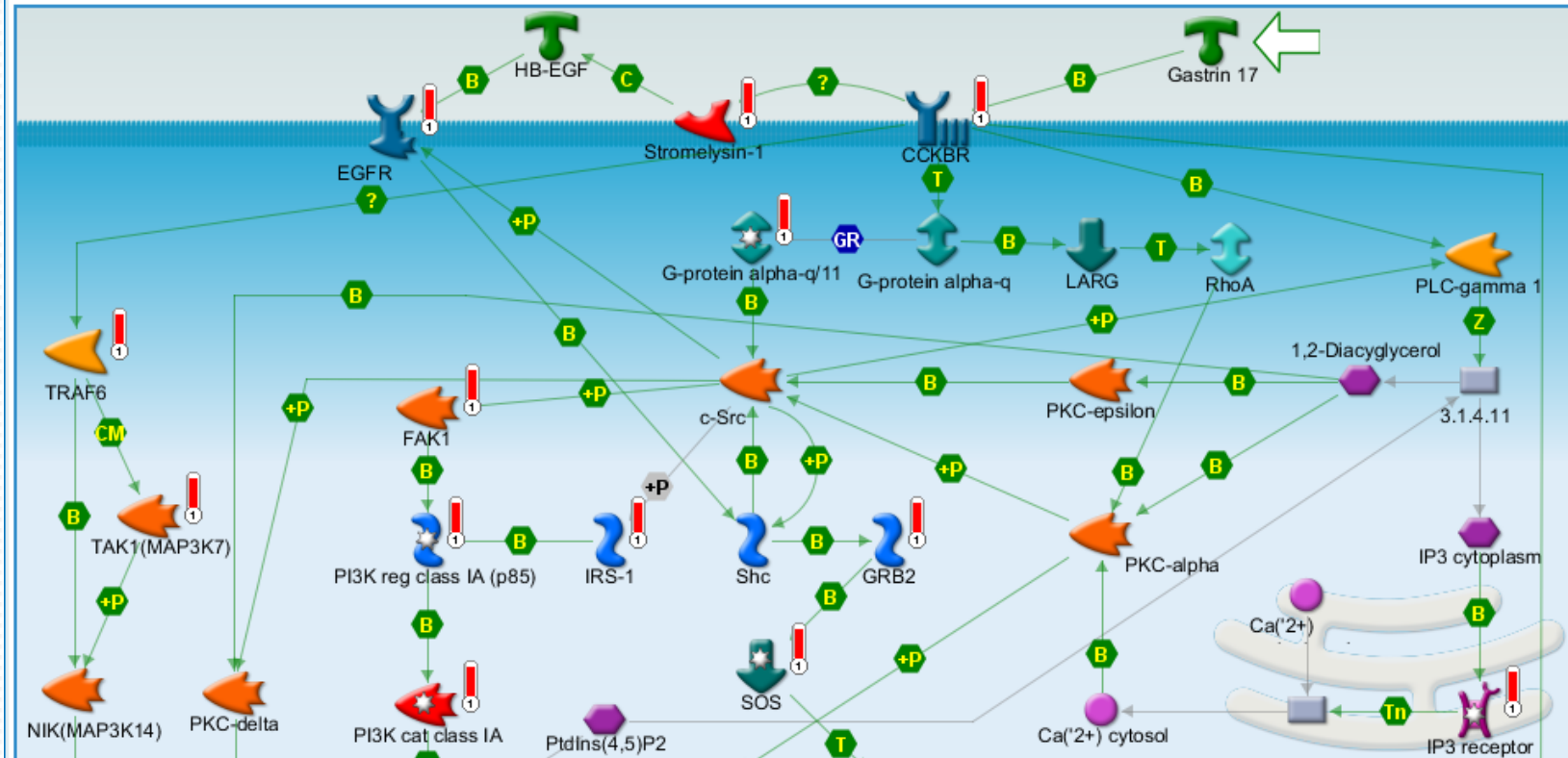
GeneGo

Immune_response_Gastrin in inflammatory response

Export to image

Experiments order

Legend



MPS - other applications

- Many new uses for this technology
- Continue to be a driver as costs go down
- Need to be careful to test and understand as the techniques may be similar but the analysis may have different assumptions/conditions

Links

- <http://wiki.g2.bx.psu.edu/Events/GCC2012/TrainingDay/VMs> -- galaxy on your laptop
- <http://trinityrnaseq.sourceforge.net/> -- trinity
- <http://www.stat.wisc.edu/~keles/Software/mosaics/> -- chip seq w/ mosaics
- <http://www.bioinformatics.babraham.ac.uk/projects/bismark/> -- methylation
- <http://sourceforge.net/p/krona/home/krona/> -- krona metagenomics display

Discussion

- Proposed projects??
 - Feedback, discussion, experiences